

**Figure 1.9** Exploded view of the main components of an ElFFF channel. *Source:* Tri et al. (2000) / American Chemical Society.

The retention parameter in ElFFF is given by

$$\lambda = \frac{D}{\mu_e E_{\text{eff}} w} = \frac{kT}{3\pi\eta d_h \mu_e E_{\text{eff}} w} \quad (1.36)$$

where  $\mu_e$  is the particle electrophoretic mobility and  $E_{\text{eff}}$  is the effective electric field (potential gradient), and it was noted that the particle hydrodynamic diameter includes the double layer thickness which is significant at low ionic strength.

Palkar and Schure (1997a) studied the time dependence of the electrode polarization effect as well as the influence of flow rate. They also studied the influence of sample size on retention and the effect of sample conductivity (Palkar and Schure 1997b). It is clear that the precise prediction of retention times in ElFFF is not a simple matter.

Micro-machined ElFFF channels were introduced in 1998 by Gale et al. (1998). The channels were 4–6 cm long, just 20–30  $\mu\text{m}$  thick, and 0.4–8 mm broad. Titanium followed by gold was sputtered onto the silicon wafer and glass plate walls to serve as electrodes (titanium was used for its good adhesion to silicon). Sample volumes were typically as small as 0.1  $\mu\text{L}$ , injected through a septum close to the channel inlet. The advantages of miniaturization of ElFFF were later examined in theory and practice (Gale et al. 2001, 2002). The advantages lie in improved efficiency, faster sample relaxation, reduced steric inversion diameter, and reduced system time constant (the time for the system to stabilize on applying the potential gradient). The reduced time constant opened up the possibility of using alternating electric fields for cyclical operation. The electrode polarization problem could then be circumvented using a square-wave, cyclical electric field with optimized frequency (Lao et al. 2002; Gale and Srinivas 2005). In cyclical operation, species are separated according to differences in their electrophoretic mobilities  $\mu_e$ , rather than in the ratios of  $D/\mu_e$ . It is the mobility and the frequency of the cycling field that determines the fraction of time spent in the faster-flowing regions as compared to the regions close to the walls (Giddings 1986; Lee et al. 1988; Stevens 1990). Cyclical ElFFF

has been shown to be an effective separation technique for submicron-charged particles. Improvements to modeling (Chen and Chauhan 2007; Kantak et al. 2006), experimental optimization (Gigault et al. 2011; Srinivas et al. 2010), and the advantage of operation with biased fields (Ornthai et al. 2015; Tasci et al. 2013) have yielded significant improvements.

## 1.8 Magnetic Field-Flow Fractionation

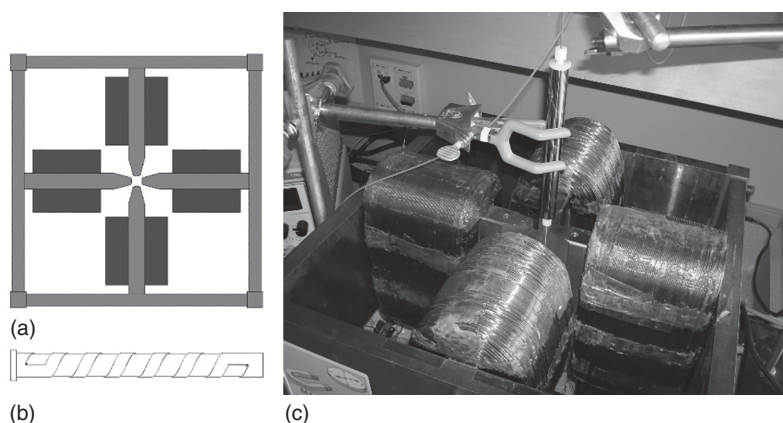
Different approaches to implementing magnetic FFF (MgFFF) have been considered through the years. Many of the early efforts were reviewed by Carpino et al. (2005b). Studies involving simple capillary tubes with transverse applied magnetic field gradients were reported by Mori (1986), Latham et al. (2005), and Vickrey and Garcia-Ramirez (1980). Tubular channel geometry with a transverse applied field is not well suited to the implementation of FFF (Giddings 2000), and the field gradients were relatively small. Mori (1986) demonstrated only slight retention of  $\text{Ni}^{2+}$  protein complexes, and Latham et al. (2005) obtained separation of 13-nm  $\text{CoFe}_2\text{O}_4$  particles in hexane from 6-nm  $\text{Fe}_2\text{O}_3$  particles that eluted with the void peak. Nomizu et al. (2001) used an intermittent transverse magnetic gradient, provided by an electromagnet, applied to a capillary tube to show separation between retained 0.6- $\mu\text{m}$  magnetite particles from effectively non-retained 0.7- $\mu\text{m}$  hematite particles in an aqueous carrier with 0.1% sodium oleate as a dispersive agent. It is not possible to determine whether retention of magnetite particles was in accord with magnetophoretic mobility as would be expected for the cyclical FFF mechanism, or whether it was simply a function of the time captured during periods of applied transverse field gradient.

Fukui, Ohara, and coworkers (Fukui et al. 2008, 2009; Takahashi et al. 2006) proposed the use of high-temperature superconducting magnets to obtain higher field gradients of 200 T/m or more. They carried out theoretical modeling of separation in capillaries subjected to such field gradients. They did not allow for relaxation to steady-state distributions before elution, however, and their simulations reflected separation due to differences in relaxation across the capillary cross-section. Therefore, the mechanism considered was not strictly that of FFF.

In 1984, Schunk, Gorse, and Burke (Gorse et al. 1984; Schunk et al. 1984) reported the use of a parallel-plate channel with a transverse field gradient generated by an electromagnet. They were able to separate singlet 0.8- $\mu\text{m}$ -rod-shaped iron oxide particles used in the recording industry from doublets. Again, the field gradient was rather small. As a means of creating high field gradients in a channel, Semenov and Kuznetsov (1986) proposed mounting a ferromagnetic wire at the axis of a tubular channel and magnetizing the wire with an external magnetic field. The concept is taken from high-gradient magnetic separation (HGMS) technology (Oberteuffer 1973) widely used nowadays for immunomagnetic cell separation. The small surface of the wire would serve as the accumulation wall, which would make the system susceptible to overloading. The field gradient would also tend to increase

rapidly with an approach to the wire which would tend to capture species which is the objective in HGMS. Semenov (1986) solved these problems by proposing that a uniform array of wires be embedded in one of the walls of a parallel-plate channel. The regular spacing of the wires was predicted to generate a fairly uniform field gradient in the channel. There have since been several modeling efforts and simulations of particle separations for such a design (Karki et al. 2001; Ohara et al. 1996; Ohara 1997; Ohara et al. 2000; Tsukamoto et al. 1995; Wang et al. 1997) and just one experimental implementation where there was shown to be slight retention of some transition metal salts (Mitsuhashi et al. 2002).

The most successful approach to MgFFF to date uses a quadrupole electromagnet and helical channel (Carpino et al. 2005a, 2005b, 2007; Williams 2012; Williams et al., 2009b, 2010c). A relatively small aperture (1–2 cm diameter) quadrupole electromagnet can generate uniform field gradients comparable to those found in the much bigger (10 cm diameter) superconducting quadrupoles (Takahashi et al. 2006), and these can be efficiently exploited using a helical channel mounted axisymmetrically to the field, close to the pole pieces. The helical channel has the advantage over a simple annular channel in that it is far simpler to maintain uniform thickness. It is also much easier to introduce fluid uniformly to the helical channel than to a full annular channel, and this is also true for the withdrawal of the fluid at the channel outlet. Also, the helical flow path carries all sample components through any small variations in field gradient around the annular space that would contribute to bandspreading in an annular channel. The use of an electromagnet also allows for very easy implementation of programmed decay of magnetic field gradient during sample analysis (Williams et al. 2010a). The quadrupole electromagnet and spiral channel are shown schematically in Figure 1.10 a and b, respectively, and a photograph of the system in Figure 1.10 c.



**Figure 1.10** (a) Schematic of the cross-section of the soft iron pole pieces and yoke (pale gray) and electrical coils (dark gray); (b) Schematic of spiral channel machined into Delrin™ (DuPont) rod that fits tightly into a stainless steel cylinder; *Source:* Carpino et al. (2005b) / with permission of ELSEVIER. (c) Photograph of the MgFFF system with the assembled spiral channel ready for an introduction to the quadrupole aperture.

The small deviations from parabolic of longitudinal and azimuthal velocity profiles in annular flow have been studied, as well as their influence on the retention ratio in a helical channel (Williams et al. 2009a, 2010b). However, if the channel thickness is relatively small compared to its radius of curvature, then the retention ratio and nonequilibrium bandspreading parameter are well approximated by the classical model equations (see Eqs. (1.2–1.6)).

The force  $F_m$  experienced by a magnetic particle in suspension placed in a magnetic field gradient is given by

$$F_m = V_m \Delta\chi \frac{B}{\mu_0} \nabla B = V_m M \nabla B \quad (1.37)$$

where  $V_m$  is the volume of magnetizable material contained in the particle,  $\Delta\chi$  is the difference in magnetic susceptibility between the magnetizable material and the other materials present (the fluid and the other particle components, all assumed to have small susceptibility),  $\mu_0$  is the magnetic permeability of free space,  $M$  is the magnetization of the magnetizable material in the particles at the applied magnetic field  $B$  (other materials assumed to have negligible magnetization), and  $\nabla B$  is the gradient in the magnitude of the magnetic field across the channel thickness. In an ideal quadrupole, the magnitude of the magnetic field  $B$  increases linearly with distance from the axis

$$B = \frac{r}{r_o} B_o \quad (1.38)$$

where  $r$  is the distance from the axis,  $r_o$  is the radius of the channel outer wall, and  $B_o$  is the magnitude of the field at  $r_o$ . The field gradient  $\nabla B$  across the channel thickness is therefore constant and equal to  $B_o/r_o$ . Replacing  $F_G$  in Eq. (1.24) by  $F_m$ , the retention parameter  $\lambda$  is given by

$$\lambda = \frac{kT}{F_m w} = \frac{kT r_o}{V_m M B_o w} \quad (1.39)$$

The magnetization is a function of local field  $B$  as may be seen in Eq. (1.37), where  $M = \chi B/\mu_0$ , and for paramagnetic and diamagnetic materials,  $\chi$  is constant. However, particles that are of particular interest for magnetic characterization are the superparamagnetic nanoparticles used for immunomagnetic labeling of specific biological cell types or used as drug carriers for magnetically targeted cancer treatment. These are composite particles where the iron oxide nanoparticles are coated with a biocompatible material such as dextran and which also may carry a chemotherapeutic drug and/or antibodies to a specific cell type. The magnetization  $M$  of the superparamagnetic component does not increase indefinitely with  $B$ . It approaches a saturation magnetization  $M_s$  at relatively low fields of 0.1–0.2 T. In this case, the concentration profile approaches an exponential, with  $\lambda$  given by Eq. (1.39) with  $M = M_s$ . Even at low field strength, where the magnetic material is approximately paramagnetic, the concentration profile does not deviate significantly from an exponential in the thin channels used.

For the characterization of superparamagnetic nanoparticles, it is not necessary for them to remain magnetically saturated during elution. If the full magnetization

curve ( $M$  as a function of  $B$  from zero to saturation) is known for the magnetic material included in a sample of magnetic nanoparticles, then it is possible to predict retention times as a function of  $V_m$ , or the equivalent spherical diameter  $d_m$ , for any applied field or programmed field decay. There is one caveat. Although only the magnetic component responds to the field gradient, the steric exclusion correction, if it is considered, is dependent on the overall particle size. They do tend to be rather small (less than about 200 nm), however, and the steric correction will be correspondingly small. Using the general integral approach developed for data reduction (Williams et al. 2001), it is also possible to transform an elution profile into a distribution in  $V_m$  or  $d_m$ . Regarding the prediction of fractionating power, nonequilibrium bandspreading is a function of  $D$  and therefore of overall particle size (see Eq. (1.4) for the contribution to plate height). It would be necessary to make some assumptions concerning particle size. For example, particles may be assumed to have similar sizes (determined by light scattering or AsFIFFF, perhaps) but contain differing amounts of magnetic components.

It should be pointed out that MgFFF applies to the fractionation of superparamagnetic nanoparticles only if they have a nonmagnetic coating. Without the coating, the magnetic interaction between the magnetized particles causes aggregation, and fractionation is not possible (Williams et al. 2010c).

## 1.9 Novel Techniques

### 1.9.1 Combined Fields

The combination of more than a single transverse field can sometimes be advantageous. Various combinations have been explored. Chen et al. (1988) showed that a steric (or hyperlayer) separation of supramicron polystyrene particle standards by symmetrical FIFFF could be improved if gravity was imposed in the same direction as the cross-flow. They called the technique gravity-augmented FIFFF.

Liu and Giddings (1991) reported the use of thermal-electrical FFF for the separation of submicron polystyrene particle standards in acetonitrile. The retention of the particles due to the thermal gradient could be enhanced or reduced by the application of a voltage gradient across the channel.

Ultrasound-gravitational FFF has been described (Yin et al., 2013) in which the force due to a resonant acoustic field on microparticles toward the node at the channel midpoint is opposed by gravity. No experimental results were presented, but particles of different compositions were expected to be driven to different equilibrium positions across the channel thickness and be carried to the outlet at different times. The technique would not have the capability to separate particles by size alone because forces due to both the standing acoustic wave and gravity depend on particle volume.

Johann et al. (2015) constructed an AsFIFFF instrument in which an electrical field can be applied across the channel thickness. They were able to determine electrophoretic mobilities of nanoparticles and proteins by measuring elution

times using cross-flow alone and then measuring the increase in elution times with the application of a voltage gradient. They were also able to show enhanced separation with the application of the electrical field. Further studies have been reported recently following the commercializing of this electrical AsFFFF system (also referred to as EAF4) (Choi et al. 2020; Metzger et al. 2021; Kohl et al. 2021).

The technique of dielectrophoretic FFF (DEP-FFF) requires the opposition of dielectrical force and gravity (Huang et al. 1997; Markx et al. 1997; Wang et al. 1998, 2000; Yang et al. 2000). A negative dielectrophoretic force on microparticles drives them away from an array of microelectrodes in the lower channel wall, and this is opposed by gravity if they are denser than the fluid. The dielectrophoretic force varies strongly with distance from the electrodes, while gravity exerts a constant force. Both forces vary with the volume of the particles. The equilibrium position and elution time depend on both the density and the polarizability of the particle. The technique has been found to separate various types of biological cells based on differences in their polarizability.

An interesting technique proposed by Janča and Audebert combines an electrical field with gravity to implement a type of hyperlayer FFF based on isopycnic focusing (Janča and Audebert 1993, 1994). A colloidal density modifier, such as Percoll, is added to the carrier fluid. This is driven by the electrical field to form a density gradient across the channel thickness, and the microparticles to be separated find their isopycnic equilibrium positions within this gradient under the influence of gravity. They were able to show the influence of voltage gradient on the retention ratios of different particles.

### 1.9.2 Two-Dimensional, Continuous Fractionation

There has been some development of two-dimensional FFF instruments for continuous fractionations. Giddings discussed the theoretical aspects of continuous FFF separations (Giddings 1984, 1990b). It was explained that a selective FFF separation in one direction can be combined with a field-induced migration at right angles resulting in different trajectories to different collection points for different species. The field-induced migration may or may not be selective, but if it is selective, its selectivity must differ from that of the FFF separation. The migration at right angles to the FFF separation may also be provided by a non-selective flow or bulk displacement.

A continuous steric FFF device was developed that used a planar channel whose breadth was set at an angle to the horizontal so that particles sedimented across the channel breadth as they migrated in steric mode along the channel length in the horizontal direction (Myers and Giddings 1979; Schure et al. 1985). Particles of different sizes could be collected at different outlets along the lower edge of the channel. Ivory et al. (1995) constructed a continuous SdFFF instrument that used a centrifuge rotor housing a channel with a conic cross-section. The channel was therefore at an angle to the radius. It was intended that the migration in the direction of flow and rotation was to be *via* the mechanism of normal or steric mode SdFFF and the migration across the channel breadth by sedimentation. Unfortunately, separation appeared to be disrupted by flow instabilities in the rotating channel.



Pearlstein and Shiue (1995) presented a concept for continuous FFF separation in the annular space between concentric cylinders, one of which rotates while the other is held stationary. The carrier fluid flows axially within the annulus and the sample is introduced at a fixed point on the circumference at the channel inlet. Sample species were predicted to follow different spiral paths along the annular channel to be collected at different points around the circumference of the outlet. They presented only a mathematical model of the expected separation for such a system.

Vastamäki and coworkers (Vastamäki et al. 2014, 2001, 2003, 2005) have developed a continuous two-dimensional ThFFF instrument. It makes use of radial carrier fluid flow between two circular plates, the upper of which is stationary and heated, while the lower is slowly rotated and is cooled. The carrier fluid is introduced at the center of the upper heated disk, and the sample is continuously fed into a second inlet that is a small distance from the axis of the upper disk. Sample species relax to the lower rotating disk and migrate radially outward by the mechanism of ThFFF. At the same time, they are angularly displaced by the rotation of the lower disk. Different species follow different curved paths to the circumference where they are collected at several collection ports around the edges of the disks.

There are numerous potentially useful combinations of fields, flows, as well as field and flow directions that may be exploited for separations of different species of differing size or composition. There are likely to be many interesting developments in the future.

## References

- Ahn, J.Y., Kim, K.H., Lee, J.Y. et al. (2010). Effect of asymmetrical flow field-flow fractionation channel geometry on separation efficiency. *Journal of Chromatography A* 1217 (24): 3876–3880. <https://doi.org/10.1016/j.chroma.2010.04.021>.
- Beckett, R., Nicholson, G., Hart, B.T. et al. (1988). Separation and size characterization of colloidal particles in river water by sedimentation field-flow fractionation. *Water Research* 22 (12): 1535–1545. [https://doi.org/10.1016/0043-1354\(88\)90166-2](https://doi.org/10.1016/0043-1354(88)90166-2).
- Beckett, R., Sharma, R., Andric, G. et al. (2007). Illustrating some principles of separation science through gravitational field-flow fractionation. *Journal of Chemical Education* 84 (12): 1955–1962. <https://doi.org/10.1021/ed084p1955>.
- Belgaied, J.E., Hoyos, M., and Martin, M. (1994). Velocity profiles in thermal field-flow fractionation. *Journal of Chromatography A* 678 (1): 85–96. [https://doi.org/10.1016/0021-9673\(94\)87077-2](https://doi.org/10.1016/0021-9673(94)87077-2).
- Berg, H.C. and Purcell, E.M. (1967). A method for separating according to mass a mixture of macromolecules or small particles suspended in a fluid, III. Experiments in a centrifugal field. *Proceedings of the National Academy of Sciences* 58 (5): 1821–1828. <https://doi.org/10.1073/pnas.58.5.1821>.
- Berg, H.C., Purcell, E.M., and Stewart, W.W. (1967). A method for separating according to mass a mixture of macromolecules or small particles suspended in a fluid, II. Experiments in a gravitational field. *Proceedings of the National Academy of Sciences* 58 (4): 1286–1291. <https://doi.org/10.1073/pnas.58.4.1286>.

- Brimhall, S.L., Myers, M.N., Caldwell, K.D., and Giddings, J.C. (1985). Study of temperature dependence of thermal diffusion in polystyrene/ethylbenzene by thermal field-flow fractionation. *Journal of Polymer Science Polymer Physics Edition* 23 (12): 2443–2456. <https://doi.org/10.1002/pol.1985.180231203>.
- Caldwell, K.D., Kesner, L.F., Myers, M.N., and Giddings, J.C. (1972). Electrical field-flow fractionation of proteins. *Science* 176 (4032): 296–298. <https://doi.org/10.1126/science.176.4032.296>.
- Caldwell, K.D., Nguyen, T.T., Myers, M.N., and Giddings, J.C. (1979). Observations on anomalous retention in steric field-flow fractionation. *Separation Science and Technology* 14 (10): 935–946. <https://doi.org/10.1080/01496397908058103>.
- Caldwell, K.D. and Gao, Y.-S. (1993). Electrical field-flow fractionation in particle separation. 1. Monodisperse standards. *Analytical Chemistry* 65 (13): 1764–1772. <https://doi.org/10.1021/ac00061a021>.
- Cao, W., Williams, P.S., Myers, M.N., and Giddings, J.C. (1999). Thermal field-flow fractionation universal calibration: extension for consideration of variation of cold wall temperature. *Analytical Chemistry* 71 (8): 1597–1608. <https://doi.org/10.1021/ac981094m>.
- Carlshaf, A. and Jönsson, J.Å. (1993). Properties of hollow fibers used for flow field-flow fractionation. *Separation Science and Technology* 28 (4): 1031–1042. <https://doi.org/10.1080/01496399308029236>.
- Carpino, F., Moore, L.R., Chalmers, J.J. et al. (2005a). Quadrupole magnetic field-flow fractionation for the analysis of magnetic nanoparticles. *Journal of Physics: Conference Series* 17: 174–180. <https://doi.org/10.1088/1742-6596/17/1/024>.
- Carpino, F., Moore, L.R., Zborowski, M. et al. (2005b). Analysis of magnetic nanoparticles using quadrupole magnetic field-flow fractionation. *Journal of Magnetism and Magnetic Materials* 293 (1): 546–552. <https://doi.org/10.1016/j.jmmm.2005.01.071>.
- Carpino, F., Zborowski, M., and Williams, P.S. (2007). Quadrupole magnetic field-flow fractionation: a novel technique for the characterization of magnetic nanoparticles. *Journal of Magnetism and Magnetic Materials* 311 (1): 383–387. <https://doi.org/10.1016/j.jmmm.2006.11.162>.
- Chen, X., Wahlund, K.-G., and Giddings, J.C. (1988). Gravity-augmented high-speed flow/steric field-flow fractionation: simultaneous use of two fields. *Analytical Chemistry* 60 (4): 362–364. <https://doi.org/10.1021/ac00155a019>.
- Chen, Z. and Chauhan, A. (2007). Electrochemical response and separation in cyclic electric field-flow fractionation. *Electrophoresis* 28 (5): 724–739. <https://doi.org/10.1002/elps.200600324>.
- Chiang, A.S., Kmietek, E.H., Langan, S.M. et al. (1979). Preliminary experimental survey of hollow-fiber electro-polarization chromatography (electrical field-flow fractionation) for protein fractionation. *Separation Science and Technology* 14 (6): 453–474. <https://doi.org/10.1080/01496397908068470>.
- Chmelik, J. (1999). Different elution modes and field programming in gravitational field-flow fractionation. I. A theoretical approach. *Journal of Chromatography A* 845 (1–2): 285–291. [https://doi.org/10.1016/S0021-9673\(99\)00131-4](https://doi.org/10.1016/S0021-9673(99)00131-4).



- Choi, J., Fuentes, C., Fransson, J. et al. (2020). Separation and zeta-potential determination of proteins and their oligomers using electrical asymmetrical flow field-flow fractionation (EAF4). *Journal of Chromatography A* 461625: <https://doi.org/10.1016/j.chroma.2020.461625>.
- Davis, J.M., Fan, F.-R.F., and Bard, A.J. (1987). Retention by electrical field-flow fractionation of anions in a new apparatus with annular porous glass channels. *Analytical Chemistry* 59 (9): 1339–1348. <https://doi.org/10.1021/ac00136a017>.
- Doshi, M.R., Gill, W.N., and Subramanian, R.S. (1975). Unsteady reverse osmosis or ultrafiltration in a tube. *Chemical Engineering Science* 30 (12): 1467–1476. [https://doi.org/10.1016/0009-2509\(75\)85024-X](https://doi.org/10.1016/0009-2509(75)85024-X).
- Fuentes, C., Choi, J., Zielke, C. et al. (2019). Comparison between conventional and frit-inlet channel in separation of biopolymers by asymmetric flow field-flow fractionation. *Analyst* 144 (15): 4559–4568. <https://doi.org/10.1039/C9AN00466A>.
- Fukui, S., Shoji, Y., Abe, R. et al. (2008). Numerical simulation of flow fractionation characteristics of magnetic chromatography using an HTS bulk magnet. *IEEE Transactions on Applied Superconductivity* 18 (2): 828–831. <https://doi.org/10.1109/TASC.2008.921248>.
- Fukui, S., Shoji, Y., Ogawa, J. et al. (2009). Study of flow fractionation characteristics of magnetic chromatography utilizing high-temperature superconducting bulk magnet. *Science and Technology of Advanced Materials* 10 (1): 014610. <https://doi.org/10.1088/1468-6996/10/1/014610>.
- Gale, B.K., Caldwell, K.D., and Frazier, A.B. (1998). A micromachined electrical field-flow fractionation ( $\mu$ -EFFF) system. *IEEE Transactions on Biomedical Engineering* 45 (12): 1459–1469. <https://doi.org/10.1109/10.730439>.
- Gale, B.K., Caldwell, K.D., and Frazier, A.B. (2001). Geometric scaling effects in electrical field flow fractionation. 1. Theoretical analysis. *Analytical Chemistry* 73 (10): 2345–2352. <https://doi.org/10.1021/ac001463q>.
- Gale, B.K., Caldwell, K.D., and Frazier, A.B. (2002). Geometric scaling effects in electrical field flow fractionation. 2. Experimental results. *Analytical Chemistry* 74 (5): 1024–1030. <https://doi.org/10.1021/ac015623p>.
- Gale, B.K. and Srinivas, M. (2005). Cyclical electrical field flow fractionation. *Electrophoresis* 26 (9): 1623–1632. <https://doi.org/10.1002/elps.200410296>.
- Giddings, J.C. (1968). Nonequilibrium theory of field-flow fractionation. *Journal of Chemical Physics* 49 (1): 81–85. <https://doi.org/10.1063/1.1669863>.
- Giddings, J.C., Yang, F.J.F., and Myers, M.N. (1974). Sedimentation field-flow fractionation. *Analytical Chemistry* 46 (13): 1917–1924. <https://doi.org/10.1021/ac60349a046>.
- Giddings, J.C., Yoon, Y.H., Caldwell, K.D. et al. (1975). Nonequilibrium plate height for field-flow fractionation in ideal parallel plate columns. *Separation Science* 10 (4): 447–460. <https://doi.org/10.1080/00372367508058032>.
- Giddings, J.C., Caldwell, K.D., and Myers, M.N. (1976a). Thermal diffusion of polystyrene in eight solvents by an improved thermal field-flow fractionation methodology. *Macromolecules* 9 (1): 106–112. <https://doi.org/10.1021/ma60049a021>.

- Giddings, J.C., Lin, G.-C., and Myers, M.N. (1976b). Electrical field-flow fractionation in a rigid membrane channel. *Separation Science* 11 (6): 553–568. <https://doi.org/10.1080/01496397608085344>.
- Giddings, J.C., Yang, F.J., and Myers, M.N. (1976c). Theoretical and experimental characterization of flow field-flow fractionation. *Analytical Chemistry* 48 (8): 1126–1132. <https://doi.org/10.1021/ac50002a016>.
- Giddings, J.C., Yang, F.J.F., and Myers, M.N. (1976d). Flow field-flow fractionation: a versatile new separation method. *Science* 193 (4259): 1244–1245. <https://doi.org/10.1126/science.959835>.
- Giddings, J.C. (1978). Displacement and dispersion of particles of finite size in flow channels with lateral forces. Field-flow fractionation and hydrodynamic chromatography. *Separation Science and Technology* 13 (3): 241–254. <https://doi.org/10.1080/01496397808060222>.
- Giddings, J.C. and Myers, M.N. (1978). Steric field-flow fractionation: a new method for separating 1 to 100  $\mu\text{m}$  particles. *Separation Science and Technology* 13 (8): 637–645. <https://doi.org/10.1080/01496397808057119>.
- Giddings, J.C. (1979). ERRATA. Displacement and dispersion of particles of finite size in flow channels with lateral forces. Field-flow fractionation and hydrodynamic chromatography. *Separation Science and Technology* 14 (9): 869–870. <https://doi.org/10.1080/01496397908060246>.
- Giddings, J.C., Myers, M.N., Caldwell, K.D., and Pav, J.W. (1979). Steric field-flow fractionation as a tool for the size characterization of chromatographic supports. *Journal of Chromatography* 185: 261–271. [https://doi.org/10.1016/S0021-9673\(00\)85608-3](https://doi.org/10.1016/S0021-9673(00)85608-3).
- Giddings, J.C. (1984). Two-dimensional separations: concept and promise. *Analytical Chemistry* 56 (12): 1258A–1264A. <https://doi.org/10.1021/ac00276a003>.
- Giddings, J.C. (1985). Optimized field-flow fractionation system based on dual stream splitters. *Analytical Chemistry* 57 (4): 945–947. <https://doi.org/10.1021/ac00281a037>.
- Giddings, J.C. (1986). Cyclical field-flow fractionation: a new method based on transport rates. *Analytical Chemistry* 58 (9): 2052–2056. <https://doi.org/10.1021/ac00122a027>.
- Giddings, J.C., Williams, P.S., and Beckett, R. (1987). Fractionating power in programmed field-flow fractionation: exponential sedimentation field decay. *Analytical Chemistry* 59 (1): 28–37. <https://doi.org/10.1021/ac00128a007>.
- Giddings, J.C. (1990a). Hydrodynamic relaxation and sample concentration in field-flow fractionation using permeable wall elements. *Analytical Chemistry* 62 (21): 2306–2312. <https://doi.org/10.1021/ac00220a010>.
- Giddings, J.C. (1990b). Two-dimensional field-flow fractionation. *Journal of Chromatography* 504: 247–258. [https://doi.org/10.1016/S0021-9673\(01\)89530-3](https://doi.org/10.1016/S0021-9673(01)89530-3).
- Giddings, J.C., Kumar, V., Williams, P.S., and Myers, M.N. (1990). Polymer separation by thermal field-flow fractionation: high-speed power programming. In: *Polymer Characterization: Physical, Spectroscopic, and Chromatographic Methods*, vol. 227, Ch. 1 (ed. C.D. Craver and T. Provder), 3–21. Washington, DC: American Chemical Society <https://doi.org/10.1021/ba-1990-0227.ch001>.

- Giddings, J.C., Moon, M.H., Williams, P.S., and Myers, M.N. (1991). Particle size distribution by sedimentation/steric field-flow fractionation: development of a calibration procedure based on density compensation. *Analytical Chemistry* 63 (14): 1366–1372. <https://doi.org/10.1021/ac00014a006>.
- Giddings, J.C. (1994). Universal calibration in size exclusion chromatography and thermal field-flow fractionation. *Analytical Chemistry* 66 (17): 2783–2787. <https://doi.org/10.1021/ac00089a029>.
- Giddings, J.C. (2000). The field-flow fractionation family: underlying principles. In: *Field-Flow Fractionation Handbook*, vol. Ch. 1 (ed. M.E. Schimpf, K. Caldwell, and J.C. Giddings), 3–30. New York, NY: Wiley.
- Gigault, J., Gale, B.K., Le Hecho, I., and Lespes, G. (2011). Nanoparticle characterization by cyclical electrical field-flow fractionation. *Analytical Chemistry* 83 (17): 6565–6572. <https://doi.org/10.1021/ac2008948>.
- Gopalakrishnan, A., Bouby, M., and Schäfer, A.I. (2023). Membrane-organic solute interactions in asymmetric flow field flow fractionation: interplay of hydrodynamic and electrostatic forces. *Science of the Total Environment* 855: 158891. <https://doi.org/10.1016/j.scitotenv.2022.158891>.
- Gorse, J., Schunk, T.C., and Burke, M.F. (1984). The study of liquid suspensions of iron oxide particles with a magnetic field-flow fractionation device. *Separation Science and Technology* 19 (13–15): 1073–1085. <https://doi.org/10.1080/01496398408058349>.
- Granger, J., Dodds, J., Leclerc, D., and Midoux, N. (1986). Flow and diffusion of particles in a channel with one porous wall: polarization chromatography. *Chemical Engineering Science* 41 (12): 3119–3128. [https://doi.org/10.1016/0009-2509\(86\)85049-7](https://doi.org/10.1016/0009-2509(86)85049-7).
- Gunderson, J.J., Caldwell, K.D., and Giddings, J.C. (1984). Influence of temperature gradients on velocity profiles and separation parameters in thermal field-flow fractionation. *Separation Science and Technology* 19 (10): 667–683. <https://doi.org/10.1080/01496398408060668>.
- Hansen, M.E. and Giddings, J.C. (1989). Retention perturbations due to particle-wall interactions in sedimentation field-flow fractionation. *Analytical Chemistry* 61 (8): 811–819. <https://doi.org/10.1021/ac00183a006>.
- Hansen, M.E., Giddings, J.C., and Beckett, R. (1989). Colloid characterization by sedimentation field-flow fractionation. VI. Perturbations due to overloading and electrostatic repulsion. *Journal of Colloid and Interface Science* 132 (2): 300–312. [https://doi.org/10.1016/0021-9797\(89\)90245-2](https://doi.org/10.1016/0021-9797(89)90245-2).
- Huang, Y., Wang, X.-B., Becker, F.F., and Gascoyne, P.R.C. (1997). Introducing dielectrophoresis as a new force field for field-flow fractionation. *Biophysical Journal* 72 (2): 1118–1129. [https://doi.org/10.1016/S0006-3495\(97\)78144-X](https://doi.org/10.1016/S0006-3495(97)78144-X).
- Ivory, C.F., Gilmartin, M., Gobie, W.A. et al. (1995). A hybrid centrifuge rotor for continuous bioprocessing. *Biotechnology Progress* 11 (1): 21–32. <https://doi.org/10.1021/bp00031a003>.
- Janča, J. and Audebert, R. (1993). New concept in focusing field-flow fractionation and thin layer isopycnic focusing: coupling of primary electric field with secondary gravitational force. *Mikrochimica Acta* 111 (4–6): 163–175. <https://doi.org/10.1007/BF01245303>.

- Janča, J. and Audebert, R. (1994). Experimental study of isopycnic focusing generated by coupled electric and gravitational field forces: use in thin layer focusing and focusing field-flow fractionation. *Mikrochimica Acta* 113 (3–6): 299–311. <https://doi.org/10.1007/BF01243620>.
- Janča, J. (2002a). Micro-channel thermal field-flow fractionation: analysis of ultra-high molar mass polymers and colloidal particles with constant and programmed field force operation. *Journal of Liquid Chromatography & Related Technologies* 25 (13–15): 2173–2191. <https://doi.org/10.1081/JLC-120014000>.
- Janča, J. (2002b). Micro-channel thermal field-flow fractionation: new challenge in analysis of macromolecules and particles. *Journal of Liquid Chromatography and Related Technologies* 25 (5): 683–704. <https://doi.org/10.1081/JLC-120003028>.
- Janča, J., Berneron, J.-F., and Boutin, R. (2003). Micro-thermal field-flow fractionation: new high-performance method for particle size distribution analysis. *Journal of Colloid and Interface Science* 260 (2): 317–323. [https://doi.org/10.1016/S0021-9797\(02\)00216-3](https://doi.org/10.1016/S0021-9797(02)00216-3).
- Janča, J. (2008). *Microthermal Field-Flow Fractionation: Analysis of Synthetic, Natural, and Biological Macromolecules and Particles*. New York: HNB Publishing.
- Johann, C., Elsenberg, S., Schuch, H., and Rösch, U. (2015). A novel instrument and method to determine the electrophoretic mobility of nanoparticles and proteins by combining electrical and flow field-flow fractionation. *Analytical Chemistry* 87 (8): 4292–4298. <https://doi.org/10.1021/ac504712n>.
- Kantak, A., Merugu, S., and Gale, B.K. (2006). Improved theory of cyclical electrical field flow fractionation. *Electrophoresis* 27 (14): 2833–2843. <https://doi.org/10.1002/elps.200500831>.
- Karki, K.C., Whitby, E.R., Patankar, S.V. et al. (2001). A numerical model for magnetic chromatography. *Applied Mathematical Modelling* 25 (5): 355–373. [https://doi.org/10.1016/S0307-904X\(00\)00057-3](https://doi.org/10.1016/S0307-904X(00)00057-3).
- Kato, H., Nakamura, A., and Banno, H. (2019). Determination of number-based size distribution of silica particles using centrifugal field-flow fractionation. *Journal of Chromatography A* 1602: 409–418. <https://doi.org/10.1016/j.chroma.2019.05.055>.
- Kesner, L.F., Caldwell, K.D., Myers, M.N., and Giddings, J.C. (1976). Performance characteristics of electrical field-flow fractionation in a flexible membrane channel. *Analytical Chemistry* 48 (13): 1834–1839. <https://doi.org/10.1021/ac50007a007>.
- Kim, Y.B., Yang, J.S., and Moon, M.H. (2018). Investigation of steric transition with field programming in frit inlet asymmetrical flow field-flow fractionation. *Journal of Chromatography A* 1576: 131–136. <https://doi.org/10.1016/j.chroma.2018.09.036>.
- Kohl, Y., Hesler, M., Drexel, R. et al. (2021). Influence of physicochemical characteristics and stability of gold and silver nanoparticles on biological effects and translocation across an intestinal barrier – a case study from in vitro to in silico. *Nanomaterials* 11 (6): 1358. <https://doi.org/10.3390/nano11061358>.
- Lao, A.I.K., Trau, D., and Hsing, I.-M. (2002). Miniaturized flow fractionation device assisted by a pulsed electric field for nanoparticle separation. *Analytical Chemistry* 74 (20): 5364–5369. <https://doi.org/10.1021/ac0257647>.

- Latham, A.H., Freitas, R.S., Schiffer, P., and Williams, M.E. (2005). Capillary magnetic field flow fractionation and analysis of magnetic nanoparticles. *Analytical Chemistry* 77 (15): 5055–5062. <https://doi.org/10.1021/ac050611f>.
- Lee, H.-L., Reis, J.F.G., Dohner, J., and Lightfoot, E.N. (1974). Single-phase chromatography: solute retardation by ultrafiltration and electrophoresis. *AIChE Journal* 20 (4): 776–784. <https://doi.org/10.1002/aic.690200420>.
- Lee, H., Kim, H., and Moon, M.H. (2005). Field programming in frit inlet asymmetrical flow field-flow fractionation/multiangle light scattering: application to sodium hyaluronate. *Journal of Chromatography A* 1089 (1–2): 203–210. <https://doi.org/10.1016/j.chroma.2005.06.069>.
- Lee, H.L. and Lightfoot, E.N. (1976). Preliminary report on ultrafiltration-induced polarization chromatography – an analog of field-flow fractionation. *Separation Science* 11 (5): 417–440. <https://doi.org/10.1080/01496397608085333>.
- Lee, S., Myers, M.N., Beckett, R., and Giddings, J.C. (1988). Particle separation and characterization by sedimentation/cyclical-field flow fractionation. *Analytical Chemistry* 60 (11): 1129–1135. <https://doi.org/10.1021/ac00162a009>.
- Lee, S., Myers, M.N., and Giddings, J.C. (1989). Hydrodynamic relaxation using stopless flow injection in split inlet sedimentation field-flow fractionation. *Analytical Chemistry* 61 (21): 2439–2444. <https://doi.org/10.1021/ac00196a023>.
- Lee, W.J., Min, B.-R., and Moon, M.H. (1999). Improvement in particle separation by hollow fiber flow field-flow fractionation and the potential use in obtaining particle size distribution. *Analytical Chemistry* 71 (16): 3446–3452. <https://doi.org/10.1021/ac981204p>.
- Lightfoot, E.N., Noble, P.T., Chiang, A.S., and Ugolini, T.A. (1981). Characterization of an improved electropolarization chromatographic system using homogenous proteins. *Separation Science and Technology* 16 (6): 619–656. <https://doi.org/10.1080/01496398108058120>.
- Litzén, A. and Wahlund, K.-G. (1991). Zone broadening and dilution in rectangular and trapezoidal asymmetrical flow field-flow fractionation channels. *Analytical Chemistry* 63 (10): 1001–1007. <https://doi.org/10.1021/ac00010a013>.
- Litzén, A. (1993). Separation speed, retention, and dispersion in asymmetrical flow field-flow fractionation as functions of channel dimensions and flow rates. *Analytical Chemistry* 65 (4): 461–470. <https://doi.org/10.1021/ac00052a025>.
- Litzén, A., Walter, J.K., Krischollek, H., and Wahlund, K.-G. (1993). Separation and quantitation of monoclonal antibody aggregates by asymmetrical flow field-flow fractionation and comparison to gel permeation chromatography. *Analytical Biochemistry* 212 (2): 469–480. <https://doi.org/10.1006/abio.1993.1356>.
- Liu, G. and Giddings, J.C. (1991). Separation of particles in nonaqueous suspensions by thermal-electrical field-flow fractionation. *Analytical Chemistry* 63 (3): 296–299. <https://doi.org/10.1021/ac00003a021>.
- Liu, G. and Giddings, J.C. (1992). Separation of particles in aqueous suspensions by thermal field-flow fractionation. Measurement of thermal diffusion coefficients. *Chromatographia* 34 (9–10): 483–492. <https://doi.org/10.1007/BF02290241>.

- Liu, M.-K., Williams, P.S., Myers, M.N., and Giddings, J.C. (1991). Hydrodynamic relaxation in flow field-flow fractionation using both split and frit inlets. *Analytical Chemistry* 63 (19): 2115–2122. <https://doi.org/10.1021/ac00019a010>.
- Markx, G.H., Rousselet, J., and Pethig, R. (1997). DEP-FFF: field-flow fractionation using non-uniform electric fields. *Journal of Liquid Chromatography & Related Technologies* 20 (16–17): 2857–2872. <https://doi.org/10.1080/10826079708005597>.
- Martin, M. and Giddings, J.C. (1981). Retention and nonequilibrium peak broadening for a generalized flow profile in field-flow fractionation. *Journal of Physical Chemistry* 85 (6): 727–733. <https://doi.org/10.1021/j150606a025>.
- Martin, M. (1996). Relative velocity profile and flow-rate in sedimentation field-flow fractionation. *Journal of High Resolution Chromatography & Chromatography Communications* 19 (9): 481–484. <https://doi.org/10.1002/jhrc.1240190902>.
- Martin, M. (1997). Time-based retention ratio for curved separation channels: application to sedimentation field-flow fractionation. *Journal of Microcolumn Separations* 9 (3): 225–232. [https://doi.org/10.1002/\(SICI\)1520-667X\(1997\)9:3<225::AID-MCS11>3.0.CO;2-7](https://doi.org/10.1002/(SICI)1520-667X(1997)9:3<225::AID-MCS11>3.0.CO;2-7).
- Martin, M. (1998). Theory of field-flow fractionation. In: *Advances in Chromatography*, vol. 39, Ch. 1 (ed. P.R. Brown and E. Grusha), 1–138. New York: Marcel Dekker Inc.
- Martin, M., van Batten, C., and Hoyos, M. (2002). Determination of thermodiffusion parameters from thermal field-flow fractionation retention data. In: *Thermal Nonequilibrium Phenomena in Liquid Mixtures*, vol. 584, Ch. 13 (ed. W. Köhler and S. Wiegand), 250–284. Berlin, Heidelberg: Springer [https://doi.org/10.1007/3-540-45791-7\\_13](https://doi.org/10.1007/3-540-45791-7_13).
- Martin, M. and Hoyos, M. (2011). On the no-field method for void time determination in flow field-flow fractionation. *Journal of Chromatography A* 1218 (27): 4117–4125. <https://doi.org/10.1016/j.chroma.2011.01.010>.
- Martin, M. and Williams, P.S. (1992). Theoretical basis of field-flow fractionation. In: *Theoretical Advancement in Chromatography and Related Techniques, NATO ASI Series C: Mathematical and Physical Sciences*, vol. 383 (ed. F. Dondi and G. Guiochon), 513–580. Dordrecht, The Netherlands: Kluwer Academic Publisher [https://doi.org/10.1007/978-94-011-2686-1\\_18](https://doi.org/10.1007/978-94-011-2686-1_18).
- Messaoud, F.A., Sanderson, R.D., Runyon, J.R. et al. (2009). An overview on field-flow fractionation techniques and their applications in the separation and characterization of polymers. *Progress in Polymer Science* 34 (4): 351–368. <https://doi.org/10.1016/j.progpolymsci.2008.11.001>.
- Metzger, C., Drexel, R., Meier, F., and Briesen, H. (2021). Effect of ultrasonication on the size distribution and stability of cellulose nanocrystals in suspension: an asymmetrical flow field-flow fractionation study. *Cellulose* 28: 10221–10238. <https://doi.org/10.1007/s10570-021-04172-3>.
- Min, B.R., Kim, S.J., Ahn, K.-H., and Moon, M.H. (2002). Hyperlayer separation in hollow fiber flow field-flow fractionation: effect of membrane materials on resolution and selectivity. *Journal of Chromatography A* 950 (1–2): 175–182. [https://doi.org/10.1016/S0021-9673\(02\)00029-8](https://doi.org/10.1016/S0021-9673(02)00029-8).



- Mitsuhashi, K., Yoshizaki, R., Ohara, T. et al. (2002). Retention of ions in a magnetic chromatograph using high-intensity and high-gradient magnetic fields. *Separation Science and Technology* 37 (16): 3635–3645. <https://doi.org/10.1081/SS-120014810>.
- Moon, M.H. (1995). Effect of carrier solutions on particle retention in flow field-flow fractionation. *Bulletin of the Korean Chemical Society* 16 (7): 613–619. <https://doi.org/10.5012/bkcs.1995.16.7.613>.
- Moon, M.H., Kwon, H., and Park, I. (1997). Stopless flow injection in asymmetrical flow field-flow fractionation using a frit inlet. *Analytical Chemistry* 69 (7): 1436–1440. <https://doi.org/10.1021/ac960897b>.
- Moon, M.H., Williams, P.S., and Kwon, H. (1999). Retention and efficiency in frit-inlet asymmetrical flow field-flow fractionation. *Analytical Chemistry* 71 (14): 2657–2666. <https://doi.org/10.1021/ac990040p>.
- Moon, M.H. (2001). Frit-inlet asymmetrical flow field-flow fractionation (FI-AFIFFF): a stopless separation technique for macromolecules and nanoparticles. *Bulletin of the Korean Chemical Society* 22 (4): 337–348. <https://doi.org/10.5012/bkcs.2001.22.4.337>.
- Moon, M.H., Williams, P.S., Kang, D., and Hwang, I. (2002). Field and flow programming in frit-inlet asymmetrical flow field-flow fractionation. *Journal of Chromatography A* 955 (2): 263–272. <https://doi.org/10.1201/b11760-16>.
- Mori, S. (1986). Magnetic field-flow fractionation using capillary tubing. *Chromatographia* 21 (11): 642–644. <https://doi.org/10.1007/BF02311919>.
- Myers, M.N., Caldwell, K.D., and Giddings, J.C. (1974). A study of retention in thermal field-flow fractionation. *Separation Science* 9 (1): 47–70. <https://doi.org/10.1080/01496397408080043>.
- Myers, M.N. and Giddings, J.C. (1979). A continuous steric FFF device for the size separation of particles. *Powder Technology* 23 (1): 15–20. [https://doi.org/10.1016/0032-5910\(79\)85021-4](https://doi.org/10.1016/0032-5910(79)85021-4).
- Nickel, C., Scherer, C., Noskov, S. et al. (2021). Observation of interaction forces by investigation of the influence of eluent additives on the retention behavior of aqueous nanoparticle dispersions in asymmetrical flow field-flow fractionation. *Journal of Chromatography A* 1637: 461840. <https://doi.org/10.1016/j.chroma.2020.461840>.
- Nomizu, T., Yamamoto, K.-i., and Watanabe, M. (2001). Magnetic chromatography for magnetic fine particles using a periodically intermittent magnetic field. *Analytical Sciences* 17: i177–i180. <https://doi.org/10.14891/analscisp.17icas.0.i177.0>.
- Oberteuffer, J.A. (1973). High gradient magnetic separation. *IEEE Transactions on Magnetics* 9 (3): 303–306. <https://doi.org/10.1109/TMAG.1973.1067673>.
- Ohara, T., Mori, S., Oda, Y. et al. (1996). Feasibility of magnetic chromatography for ultra-fine particle separation. *Transactions of the IEE Japan* 116 (8): 979–986. [https://doi.org/10.1541/ieejpes1990.116.8\\_979](https://doi.org/10.1541/ieejpes1990.116.8_979).
- Ohara, T. (1997). Feasibility of using magnetic chromatography for ultra-fine particle separation. In: *High Magnetic Fields: Applications, Generations, Materials* (ed. H.J. Schneider-Muntau), 43–55. Hackensack, NJ: World Scientific.
- Ohara, T., Wang, X., Wada, H., and Whitby, E.R. (2000). Magnetic chromatography: numerical analysis in the case of particle size distribution. *Transactions of the IEE Japan* 120 (1): 62–67. [https://doi.org/10.1541/ieejfms1990.120.1\\_62](https://doi.org/10.1541/ieejfms1990.120.1_62).

- Ornthai, M., Siripinyanond, A., and Gale, B.K. (2015). Biased cyclical electrical field-flow fractionation for separation of submicron particles. *Analytical and Bioanalytical Chemistry* 408 (3): 855–863. <https://doi.org/10.1007/s00216-015-9173-5>.
- Palkar, S.A. and Schure, M.R. (1997a). Mechanistic study of electrical field flow fractionation. 1. Nature of the internal field. *Analytical Chemistry* 69 (16): 3223–3229. <https://doi.org/10.1021/ac9700134>.
- Palkar, S.A. and Schure, M.R. (1997b). Mechanistic study of electrical field flow fractionation. 2. Effect of sample conductivity on retention. *Analytical Chemistry* 69 (16): 3230–3238. <https://doi.org/10.1021/ac970014w>.
- Park, M.R., Kang, D.Y., Chmelik, J. et al. (2008). Different elution modes and field programming in gravitational field-flow fractionation: effect of channel angle. *Journal of Chromatography A* 1209 (1–2): 206–211. <https://doi.org/10.1016/j.chroma.2008.09.014>.
- Pearlstein, A.J. and Shiue, M.-P. (1995). Three-dimensional field-flow fractionation using helical flow. *Separation Science and Technology* 30 (11): 2251–2258. <https://doi.org/10.1080/01496399508013110>.
- Plocková, J. and Chmelík, J. (2000). Different elution modes and field programming in gravitational field-flow fractionation. 2. Experimental verification of the range of conditions for flow-rate and carrier liquid density programming. *Journal of Chromatography A* 868 (2): 217–227. [https://doi.org/10.1016/S0021-9673\(99\)01235-2](https://doi.org/10.1016/S0021-9673(99)01235-2).
- Plocková, J. and Chmelík, J. (2001). Different elution modes and field programming in gravitational field-flow fractionation. III. Field programming by flow-rate gradient generated by a programmable pump. *Journal of Chromatography A* 918 (2): 361–370. [https://doi.org/10.1016/S0021-9673\(01\)00706-3](https://doi.org/10.1016/S0021-9673(01)00706-3).
- Plocková, J., Matulík, F., and Chmelík, J. (2002). Different elution modes and field programming in gravitational field-flow fractionation. IV. Field programming achieved with channels of non-constant cross-sections. *Journal of Chromatography A* 955 (1): 95–103. [https://doi.org/10.1016/S0021-9673\(02\)00195-4](https://doi.org/10.1016/S0021-9673(02)00195-4).
- Plocková, J. and Chmelík, J. (2006). Different elution modes and field programming in gravitational field-flow fractionation: field programming using density and viscosity gradients. *Journal of Chromatography A* 1118 (2): 253–260. <https://doi.org/10.1016/j.chroma.2006.03.124>.
- Reis, J.F.G. and Lightfoot, E.N. (1976). Electropolarization chromatography. *AIChE Journal* 22 (4): 779–785. <https://doi.org/10.1002/aic.690220423>.
- Reis, J.F.G., Ramkrishna, D., and Lightfoot, E.N. (1978). Convective mass transfer in the presence of polarizing fields: dispersion in hollow fiber electropolarization chromatography. *AIChE Journal* 24 (4): 679–686. <https://doi.org/10.1002/aic.690240416>.
- Schimpf, M.E. and Giddings, J.C. (1989). Characterization of thermal diffusion in polymer solutions by thermal field-flow fractionation: dependence on polymer and solvent parameters. *Journal of Polymer Science: Part B: Polymer Physics* 27 (6): 1317–1332. <https://doi.org/10.1002/polb.1989.090270610>.
- Schimpf, M.E. and Wahlund, K.-G. (1997). Asymmetrical flow field-flow fractionation as a method to study the behavior of humic acids in solution. *Journal of Microcolumn Separations* 9 (7): 535–543. [https://doi.org/10.1002/\(SICI\)1520-667X\(1997\)9:7<535::AID-MCS3>3.0.CO;2-2](https://doi.org/10.1002/(SICI)1520-667X(1997)9:7<535::AID-MCS3>3.0.CO;2-2).

- Schimpf, M.E. and Semenov, S.N. (2000). Mechanism of polymer thermophoresis in nonaqueous solvents. *The Journal of Physical Chemistry B* 104 (42): 9935–9942. <https://doi.org/10.1021/jp994334q>.
- Schunk, T.C., Gorse, J., and Burke, M.F. (1984). Parameters affecting magnetic field-flow fractionation of metal oxide particles. *Separation Science and Technology* 19 (10): 653–666. <https://doi.org/10.1080/01496398408060667>.
- Schure, M.R., Myers, M.N., Caldwell, K.D. et al. (1985). Separation of coal fly ash using continuous steric field-flow fractionation. *Environmental Science and Technology* 19 (8): 686–689. <https://doi.org/10.1021/es00138a005>.
- Schure, M.R. and Weeratunga, S.K. (1991). Coriolis-induced secondary flow in sedimentation field-flow fractionation. *Analytical Chemistry* 63 (22): 2614–2626. <https://doi.org/10.1021/ac00022a015>.
- Semenov, S.N. (1986). Flow fractionation in a strong transverse magnetic field. *Russian Journal of Physical Chemistry* 60 (5): 729–731.
- Semenov, S.N. and Kuznetsov, A.A. (1986). Flow fractionation in a transverse high-gradient magnetic field. *Russian Journal of Physical Chemistry* 60 (2): 247–250.
- Semenov, S.N. and Schimpf, M.E. (2020). Comparative examination of nonequilibrium thermodynamic models of thermodiffusion in liquids. *Proceedings* 46 (1): 14. <https://doi.org/10.3390/ecea-5-06680>.
- Shah, A.B., Reis, J.F.G., Lightfoot, E.N., and Moore, R.E. (1979). Modeling electroretention of proteins during electropolarization chromatography. *Separation Science and Technology* 14 (6): 475–497. <https://doi.org/10.1080/01496397908068471>.
- Shiundu, P.M., Liu, G., and Giddings, J.C. (1995). Separation of particles in nonaqueous suspensions by thermal field-flow fractionation. *Analytical Chemistry* 67 (15): 2705–2713. <https://doi.org/10.1021/ac00111a032>.
- Shiundu, P.M., Munguti, S.M., and Williams, S.K.R. (2003). Retention behavior of metal particle dispersions in aqueous and nonaqueous carriers in thermal field-flow fractionation. *Journal of Chromatography A* 983 (1–2): 163–176. [https://doi.org/10.1016/S0021-9673\(02\)01694-1](https://doi.org/10.1016/S0021-9673(02)01694-1).
- Srinivas, M., Sant, H.J., and Gale, B.K. (2010). Optimization of cyclical electrical field flow fractionation. *Electrophoresis* 31 (20): 3372–3379. <https://doi.org/10.1002/elps.201000024>.
- Stevens, F.J. (1990). Fractionation of macromolecules in an alternating transverse electric field: simulation of the method. *Journal of Biochemical and Biophysical Methods* 20 (4): 275–292. [https://doi.org/10.1016/0165-022X\(90\)90090-Y](https://doi.org/10.1016/0165-022X(90)90090-Y).
- Takahashi, M., Fukui, S., Takahashi, Y. et al. (2006). Numerical study on magnetic chromatography using quadrupole magnetic field. *IEEE Transactions on Applied Superconductivity* 16 (2): 1116–1119. <https://doi.org/10.1109/TASC.2006.871336>.
- Tasci, T.O., Johnson, W.P., Fernandez, D.P. et al. (2013). Biased cyclical electrical field flow fractionation for separation of sub 50 nm particles. *Analytical Chemistry* 85 (23): 11225–11232. <https://doi.org/10.1021/ac401331z>.
- Thompson, G.H., Myers, M.N., and Giddings, J.C. (1967). An observation of a field-flow fractionation effect with polystyrene samples. *Separation Science* 2 (6): 797–900. <https://doi.org/10.1080/01496396708049739>.

- Thompson, G.H., Myers, M.N., and Giddings, J.C. (1969). Thermal field-flow fractionation of polystyrene samples. *Analytical Chemistry* 41 (10): 1219–1222. <https://doi.org/10.1021/ac60279a001>.
- Tri, N., Caldwell, K., and Beckett, R. (2000). Development of electrical field-flow fractionation. *Analytical Chemistry* 72 (8): 1823–1829. <https://doi.org/10.1021/ac990822i>.
- Tsukamoto, O., Ohizumi, T., Ohara, T. et al. (1995). Feasibility study on separation of several tens nanometer scale particles by magnetic field-flow-fractionation technique using superconducting magnet. *IEEE Transactions on Applied Superconductivity* 5 (2): 311–314. <https://doi.org/10.1109/77.402551>.
- van Asten, A.C., Boelens, H.F.M., Kok, W.T. et al. (1994). Temperature dependence of solvent viscosity, solvent thermal conductivity, and Soret coefficient in thermal field-flow fractionation. *Separation Science and Technology* 29 (4): 513–533. <https://doi.org/10.1080/01496399408002159>.
- Vastamäki, P., Jussila, M., and Riekkola, M.-L. (2001). Development of continuously operating two-dimensional thermal field-flow fractionation equipment. *Separation Science and Technology* 36 (11): 2535–2545. <https://doi.org/10.1081/SS-100106108>.
- Vastamäki, P., Jussila, M., and Riekkola, M.-L. (2003). Study of continuous two-dimensional thermal field-flow fractionation of polymers. *The Analyst* 128 (10): 1243–1248. <https://doi.org/10.1039/b307292b>.
- Vastamäki, P., Jussila, M., and Riekkola, M.-L. (2005). Continuous two-dimensional field-flow fractionation: a novel technique for continuous separation and collection of macromolecules and particles. *The Analyst* 130 (4): 427–432. <https://doi.org/10.1039/b410046h>.
- Vastamäki, P., Williams, P.S., Matti, J. et al. (2014). Retention in continuous two-dimensional thermal field-flow fractionation: comparison of experimental results with theory. *The Analyst* 139 (1): 116–127. <https://doi.org/10.1039/C3AN01047C>.
- Vauthier, J.-C. and Williams, P.S. (1998). Numerical simulation of band-broadening during hydrodynamic relaxation in frit-inlet field-flow fractionation channels. *Journal of Chromatography A* 805 (1–2): 149–160. [https://doi.org/10.1016/S0021-9673\(98\)00009-0](https://doi.org/10.1016/S0021-9673(98)00009-0).
- Vickrey, T.M. and Garcia-Ramirez, J.A. (1980). Magnetic field-flow fractionation: theoretical basis. *Separation Science and Technology* 15 (6): 1297–1304. <https://doi.org/10.1080/01496398008068506>.
- Wahlund, K.-G., Winegarner, H.S., Caldwell, K.D., and Giddings, J.C. (1986). Improved flow field-flow fractionation system applied to water soluble polymers: programming, outlet stream splitting, and flow optimization. *Analytical Chemistry* 58 (3): 573–578. <https://doi.org/10.1021/ac00294a018>.
- Wahlund, K.-G. and Giddings, J.C. (1987). Properties of an asymmetrical flow field-flow fractionation channel having one permeable wall. *Analytical Chemistry* 59 (9): 1332–1339. <https://doi.org/10.1021/ac00136a016>.
- Wahlund, K.-G. and Litzén, A. (1989). Application of an asymmetrical flow field-flow fractionation channel to the separation and characterization of proteins, plasmids, plasmid fragments, polysaccharides and unicellular algae. *Journal of Chromatography* 461: 73–87. [https://doi.org/10.1016/S0021-9673\(00\)94276-6](https://doi.org/10.1016/S0021-9673(00)94276-6).

- Wahlund, K.-G. (2013). Flow field-flow fractionation: critical overview. *Journal of Chromatography A* 1287: 97–112. <https://doi.org/10.1016/j.chroma.2013.02.028>.
- Wang, X.-B., Vykoukal, J., Becker, F.F., and Gascoyne, P.R.C. (1998). Separation of polystyrene microbeads using dielectrophoretic/gravitational field-flow-fractionation. *Biophysical Journal* 74 (5): 2689–2701. [https://doi.org/10.1016/S0006-3495\(98\)77975-5](https://doi.org/10.1016/S0006-3495(98)77975-5).
- Wang, X.-B., Yang, J., Huang, Y. et al. (2000). Cell separation by dielectrophoretic field-flow-fractionation. *Analytical Chemistry* 72 (4): 832–839. <https://doi.org/10.1021/ac990922o>.
- Wang, X., Ohara, T., Whitby, E.R. et al. (1997). Computer simulation of magnetic chromatography system for ultra-fine particle separation. *Transactions of the IEE Japan* 117 (11): 1466–1474. [https://doi.org/10.1541/ieejpes1990.117.11\\_1466](https://doi.org/10.1541/ieejpes1990.117.11_1466).
- Williams, P.S. and Giddings, J.C. (1987). Power programmed field-flow fractionation: a new program form for improved uniformity of fractionating power. *Analytical Chemistry* 59 (17): 2038–2044. <https://doi.org/10.1021/ac00144a007>.
- Williams, P.S., Giddings, J.C., and Beckett, R. (1987). Fractionating power in sedimentation field-flow fractionation with linear and parabolic field decay programming. *Journal of Liquid Chromatography* 10 (8–9): 1961–1998. <https://doi.org/10.1080/01483918708066808>.
- Williams, P.S., Koch, T., and Giddings, J.C. (1992). Characterization of near-wall hydrodynamic lift forces using sedimentation field-flow fractionation. *Chemical Engineering Communications* 111: 121–147. <https://doi.org/10.1080/00986449208935984>.
- Williams, P.S. and Giddings, J.C. (1994). Theory of field-programmed field-flow fractionation with corrections for steric effects. *Analytical Chemistry* 66 (23): 4215–4228. <https://doi.org/10.1021/ac00095a017>.
- Williams, P.S., Lee, S., and Giddings, J.C. (1994). Characterization of hydrodynamic lift forces by field-flow fractionation. Inertial and near-wall lift forces. *Chemical Engineering Communications* 130 (1): 143–166. <https://doi.org/10.1080/00986449408936272>.
- Williams, P.S., Moon, M.H., and Giddings, J.C. (1996a). Influence of accumulation wall and carrier solution composition on lift force in sedimentation/steric field-flow fractionation. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 113 (3): 215–228. [https://doi.org/10.1016/0927-7757\(96\)03669-2](https://doi.org/10.1016/0927-7757(96)03669-2).
- Williams, P.S., Moon, M.H., Xu, Y., and Giddings, J.C. (1996b). Effect of viscosity on retention time and hydrodynamic lift forces in sedimentation/steric field-flow fractionation. *Chemical Engineering Science* 51 (19): 4477–4488. [https://doi.org/10.1016/0009-2509\(96\)00291-6](https://doi.org/10.1016/0009-2509(96)00291-6).
- Williams, P.S. (1997). Design of an asymmetrical flow field-flow fractionation channel for uniform channel flow velocity. *Journal of Microcolumn Separations* 9 (6): 459–467. [https://doi.org/10.1002/\(SICI\)1520-667X\(1997\)9:6<459::AID-MCS3>3.0.CO;2-0](https://doi.org/10.1002/(SICI)1520-667X(1997)9:6<459::AID-MCS3>3.0.CO;2-0).
- Williams, P.S. (2022). Theoretical principles of field-flow fractionation and SPLITT fractionation. In: *Particle Separation Techniques* (ed. C. Contado), 579–620. Amsterdam, The Netherlands: Elsevier <https://doi.org/10.1016/B978-0-323-85486-3.00001-9>.

- Williams, P.S., Xu, Y., Reschiglian, P., and Giddings, J.C. (1997). Colloid characterization by sedimentation field-flow fractionation: correction for particle-wall interaction. *Analytical Chemistry* 69 (3): 349–360. <https://doi.org/10.1021/ac9606012>.
- Williams, P.S. (2000a). Programmed field-flow fractionation: fractionating power and optimization. In: *Field-Flow Fractionation Handbook*. (Ch 10 (ed. M.E. Schimpf, K. Caldwell, and J.C. Giddings), 167–182. New York, NY: Wiley-Interscience.
- Williams, P.S., Giddings, M.C., and Giddings, J.C. (2001). A data analysis algorithm for programmed field-flow fractionation. *Analytical Chemistry* 73 (17): 4202–4211. <https://doi.org/10.1021/ac010305b>.
- Williams, P.S., Carpino, F., and Zborowski, M. (2009a). Theory for nanoparticle retention time in the helical channel of quadrupole magnetic field-flow fractionation. *Journal of Magnetism and Magnetic Materials* 321 (10): 1446–1451. <https://doi.org/10.1016/j.jmmm.2009.02.065>.
- Williams, P.S., Carpino, F., and Zborowski, M. (2009b). Magnetic nanoparticle drug carriers and their study by quadrupole magnetic field-flow fractionation. *Molecular Pharmaceutics* 6 (5): 1290–1306. <https://doi.org/10.1021/mp900018v>.
- Williams, P.S., Carpino, F., Moore, L.R., and Zborowski, M. (2010a). Magnetic field programming in quadrupole magnetic field-flow fractionation. *Physics Procedia* 9: 91–95. <https://doi.org/10.1016/j.phpro.2010.11.022>.
- Williams, P.S., Carpino, F., and Zborowski, M. (2010b). Erratum to: “Theory for nanoparticle retention time in the helical channel of quadrupole magnetic field-flow fractionation” [J. Magn. Magn. Mater. 321 (2009) 1446–1451]. *Journal of Magnetism and Magnetic Materials* 322 (21): 3605. <https://doi.org/10.1016/j.jmmm.2010.07.011>.
- Williams, P.S., Carpino, F., and Zborowski, M. (2010c). Characterization of magnetic nanoparticles using programmed quadrupole magnetic field-flow fractionation. *Philosophical Transactions of the Royal Society A: Mathematical, Physical & Engineering Sciences* 368 (1927): 4419–4437. <https://doi.org/10.1098/rsta.2010.0133>.
- Williams, P.S. (2012). Separation and characterization of magnetic particulate materials. In: *Magnetic Nanoparticles: From Fabrication to Clinical Applications* (Ch. 11 (ed. N.T.K. Thanh), 301–332. Boca Raton, FL: CRC Press <https://doi.org/10.1201/b11760-16>.
- Williams, P.S. (2015). Retention ratio and nonequilibrium bandspreading in asymmetrical flow field-flow fractionation. *Analytical and Bioanalytical Chemistry* 407 (15): 4327–4338. <https://doi.org/10.1007/s00216-015-8734-y>.
- Williams, P.S. (2016). Fractionating power and outlet stream polydispersity in asymmetrical flow field-flow fractionation. Part I: isocratic operation. *Analytical and Bioanalytical Chemistry* 408 (12): 3247–3263. <https://doi.org/10.1007/s00216-016-9388-0>.
- Williams, P.S. (2017). Fractionating power and outlet stream polydispersity in asymmetrical flow field-flow fractionation. Part II: programmed operation. *Analytical and Bioanalytical Chemistry* 409 (1): 317–334. <https://doi.org/10.1007/s00216-016-0007-x>.



- Williams, P.S. (2024). Correction to: Fractionating power and outlet stream polydispersity in asymmetrical flow field-flow fractionation. Part II: programmed operation. *Analytical and Bioanalytical Chemistry* 416 (28): 6687–6689. <https://doi.org/10.1007/s00216-024-05527-y>.
- Williams, S.K.R. (2000b). Sample recovery. In: *Field-Flow Fractionation Handbook* (Ch. 21 (ed. M.E. Schimpf, K. Caldwell, and J.C. Giddings), 325–343. New York: Wiley-Interscience.
- Yang, J., Huang, Y., Wang, X.-B. et al. (2000). Differential analysis of human leukocytes by dielectrophoretic field-flow-fractionation. *Biophysical Journal* 78 (5): 2680–2689. [https://doi.org/10.1016/S0006-3495\(00\)76812-3](https://doi.org/10.1016/S0006-3495(00)76812-3).
- Yin, H.-Y., Zuo, C.-C., and Shi, H.-Y. (2013). Fabrication of ultrasound-gravity field flow fractionation devices using engraving machine and reliable sealing method. *Advanced Materials Research* 748: 765–768. <https://doi.org/10.4028/www.scientific.net/AMR.748.765>.
- You, Z., Meier, F., and Weidner, S. (2017). Comparison of miniaturized and conventional asymmetrical flow field-flow fractionation (AF4) channels for nanoparticle separations. *Separations* 4 (1): 8. <https://doi.org/10.3390/separations4010008>.
- Zheng, F. (2002). Thermophoresis of spherical and non-spherical particles: a review of theories and experiments. *Advances in Colloid and Interface Science* 97 (1–3): 255–278. [https://doi.org/10.1016/S0001-8686\(01\)00067-7](https://doi.org/10.1016/S0001-8686(01)00067-7).

